

Platelet Function Disorder Panel

Versiti offers comprehensive genetic analysis to detect sequence variants and large deletions and duplications in 41 genes known to cause platelet function disorders and/or inherited thrombocytopenia. This panel can be ordered as:

- **Next Generation Sequencing (NGS) only*;**
 - **NGS with reflex to Array Comparative Genomic Hybridization (aCGH) Deletion/Duplication if clinically significant variants explaining the patient's phenotype are not detected by sequencing;**
 - **NGS with concurrent aCGH Deletion/Duplication (both testing methodologies performed simultaneously); or**
 - **Deletion/Duplication by aCGH only.**
- * Includes *PLAU* performed by aCGH**
-

Inherited platelet function disorders are a heterogeneous group of bleeding disorders of variable severity caused by defects in platelet adhesion, glycoprotein expression, receptor function, signaling pathways, aggregation, cytoskeleton proteins, secretion, granular contents and abnormalities in procoagulant activity. Some have concomitant thrombocytopenia, typically characterized by platelet counts less than 150,000/uL, but often can vary with age, sex and ethnic background. Symptoms of platelet dysfunction may include purpura, petechiae, epistaxis, extensive bruising, menorrhagia in women, and the possibility of life-threatening excessive bleeding when challenged by surgery, injury, or childbirth. Disorders causing severe platelet dysfunction can present in the newborn period or early childhood, while mild platelet dysfunction may remain undiagnosed for many years. Some inherited platelet function disorders have only

hematologic manifestations, such as bleeding and low platelet counts, while other syndromic types present with additional non-hematologic manifestations. Certain types of inherited platelet disorders cause predisposition to myeloid neoplasms.

Misdiagnosis of platelet disorders can result in inappropriate therapies and inadequate surveillance for additional medical complications, underscoring the importance of accurate diagnosis. The diagnosis may be difficult to establish based solely on functional studies, as platelet function assays are not widely available due to their technical complexity and need for immediate testing on fresh patient platelets due to limited sample stability. Advances in genetic testing through next-generation sequencing allows for identification of underlying genetic defects and for distinguishing inherited platelet disorder cases from other acquired disorders of platelet function, such as those induced by medications or systemic disorders. Accurate diagnosis provides information about the phenotype and prognosis, guides medical management decisions, assists with the identification of affected family members, and allows for accurate genetic recurrence risk assessment. Variants in genes known to be associated syndromic or non-syndromic platelet dysfunction may be inherited in an autosomal recessive, autosomal dominant or X-linked recessive manner. More common and rare types of platelet disorders will be identified with this panel.

The NGS panel evaluates for single nucleotide variants and small deletions and duplications, which are most commonly responsible for genetic disease. However, large deletions and duplications, also referred to as copy number variations (CNVs), are a known cause of genetic disorders, but can escape detection by next generation sequence analysis. Additional testing with aCGH Deletion/Duplication analysis is available for all genes on this panel to evaluate for large deletions and duplications encompassing one or more exons, or affecting an entire gene. Note that the Quebec Platelet Disorder (QPD) is associated with a heterozygous 77.9-kb tandem duplication of the *PLAU* gene which will be detected by aCGH and not by NGS; analysis of *PLAU* by aCGH is included in the otherwise NGS-only version of this panel.



For broader evaluation of unspecified platelet problems, all genes on the Platelet Function Disorder Panel and the Inherited Thrombocytopenia Panel can be analyzed together by ordering the Comprehensive Platelet Disorder Panel. For broader evaluation of unspecified bleeding disorders, all genes on the Platelet Function Disorder Panel and Coagulation Disorder Panel can be analyzed together by ordering the Comprehensive Bleeding Disorder Panel.

Refer to the table for further information about each gene in the Platelet Function Disorder Panel, including the clinical phenotype and inheritance pattern.

Platelet Function Disorder Panel: gene, clinical phenotype and inheritance pattern		
Gene	Clinical Phenotype	Inheritance
<i>ANO6</i>	Scott syndrome: platelet dysfunction with mild to moderate bleeding phenotype with normal platelet aggregation and platelet counts, and decreased platelet procoagulant activity with characteristic flow cytometry findings	Autosomal Recessive
<i>AP3B1</i>	Hermansky-Pudlak syndrome type 2 (HPS2): oculocutaneous albinism of variable severity and mild bleeding due to a platelet storage pool disorder, as well as pulmonary fibrosis and neutropenia	Autosomal Recessive
<i>AP3D1</i>	Hermansky-Pudlak syndrome type 10 (HPS10): oculocutaneous albinism of variable severity and mild bleeding due to a platelet storage pool disorder, as well as neutropenia, seizures and developmental delay	Autosomal Recessive
<i>ARPC1B</i>	ARPC1B-related thrombocytopenia: microthrombocytopenia, decreased platelet dense granules, allergic and inflammatory disease	Autosomal Recessive
<i>BLOC1S3</i>	Hermansky-Pudlak syndrome type 8 (HPS8): oculocutaneous albinism of variable severity and mild bleeding due to a platelet storage pool disorder	Autosomal Recessive
<i>BLOC1S6</i>	Hermansky-Pudlak syndrome type 9 (HPS9): oculocutaneous albinism of variable severity and mild bleeding due to a platelet storage pool disorder	Autosomal Recessive
<i>DTNBP1</i>	Hermansky-Pudlak syndrome type 7 (HPS7): oculocutaneous albinism of variable severity and mild bleeding due to a platelet storage pool disorder	Autosomal Recessive
<i>FERMT3</i>	Leukocyte adhesion deficiency-III (LAD-III): characterized by a severe bleeding phenotype with a Glanzmann thrombasthenia-like phenotype on platelet aggregation studies and associated immunodeficiency	Autosomal Recessive
<i>FLII</i>	FLII-related thrombocytopenia (platelet-type bleeding disorder-21): macrothrombocytopenia with moderate bleeding from platelet dysfunction due to alpha granule deficiency (large/fused platelet alpha granules on platelet electron microscopy), with or without delta granule deficiency	Autosomal Dominant
<i>FLNA</i>	FLNA-related thrombocytopenia: macrothrombocytopenia with or without associated periventricular heterotopia.	X-linked
<i>FYB1</i>	FYB1-related thrombocytopenia (thrombocytopenia 3): non-syndromic microthrombocytopenia and platelet dysfunction leading to increased bleeding	Autosomal Recessive
<i>GATA1</i>	GATA1-related X-linked cytopenia: characterized by macrothrombocytopenia and/or anemia with moderate bleeding due to platelet alpha granule deficiency	X-linked
<i>GF11B</i>	GF11B-related thrombocytopenia (platelet-type bleeding disorder-17): macrothrombocytopenia with platelet alpha granule deficiency leading to variable bleeding tendency, red cell anisopoikilocytosis, increased numbers of dysplastic megakaryocytes and increased platelet CD34 expression	Autosomal Dominant
<i>GP1BA</i>	Bernard Soulier syndrome (BSS): macrothrombocytopenia with normal platelet granularity and moderate to severe bleeding due to decreased/absent/dysfunctional platelet GPIb/IX expression with decreased/absent platelet aggregation with ristocetin	Autosomal Recessive
<i>GP1BB</i>	Bernard Soulier syndrome (BSS): macrothrombocytopenia with normal platelet granularity and moderate to severe bleeding due to decreased/absent/dysfunctional platelet GPIb/IX expression with decreased/absent platelet aggregation with ristocetin.	Autosomal Recessive
<i>GP6</i>	GP6-related platelet dysfunction (platelet-type bleeding disorder 11): mild bleeding and decreased aggregation response to collagen on platelet aggregation studies due to deficiency of platelet glycoprotein VI	Autosomal Recessive
<i>GP9</i>	Bernard Soulier syndrome (BSS): macrothrombocytopenia with normal platelet granularity and moderate to severe bleeding due to decreased/absent/dysfunctional platelet GPIb/IX expression with decreased/absent platelet aggregation with ristocetin.	Autosomal Recessive
<i>HPS1</i>	Hermansky-Pudlak syndrome type 1 (HPS1): oculocutaneous albinism of variable severity and mild bleeding due to a platelet storage pool disorder, as well as pulmonary fibrosis and granulomatous colitis	Autosomal Recessive
<i>HPS3</i>	Hermansky-Pudlak syndrome type 3 (HPS3): mild ocular albinism and mild bleeding due to a platelet storage pool disorder	Autosomal Recessive
<i>HPS4</i>	Hermansky-Pudlak syndrome type 4 (HPS4): oculocutaneous albinism and mild bleeding due to a platelet storage pool disorder, as well as pulmonary fibrosis and granulomatous colitis	Autosomal Recessive

Platelet Function Disorder Panel: gene, clinical phenotype and inheritance pattern		
<i>HPS5</i>	Hermansky-Pudlak syndrome type 5 (HPS5): mild ocular albinism and mild bleeding due to a platelet storage pool disorder	Autosomal Recessive
<i>HPS6</i>	Hermansky-Pudlak syndrome type 6 (HPS6): mild ocular albinism and mild bleeding due to a platelet storage pool disorder	Autosomal Recessive
<i>ITGA2B</i>	Glanzmann thrombasthenia: normal platelet count with severe bleeding and decreased/absent platelet aggregation with all agonists except ristocetin due to decreased/absent/dysfunctional expression of platelet glycoprotein (GP) IIb/IIIa	Autosomal Recessive
<i>ITGB3</i>	Glanzmann thrombasthenia: normal platelet count with severe bleeding and decreased/absent platelet aggregation with all agonists except ristocetin due to decreased/absent/dysfunctional expression of platelet glycoprotein (GP) IIb/IIIa	Autosomal Recessive
<i>KDSR</i>	KDSR-related thrombocytopenia (Erythrokeratoderma variabilis et progressiva 4): thrombocytopenia with normal platelet size and platelet dysfunction with or without skin hyperkeratosis and ichthyosis	Autosomal Recessive
<i>LYST</i>	Chediak-Higashi syndrome: partial oculocutaneous albinism, immunodeficiency, and a mild bleeding from platelet delta granule deficiency	Autosomal Recessive
<i>NBEA</i>	NBEA-related platelet dysfunction: neurodevelopmental disorders, including autism and seizures and moderate bleeding due to platelet delta storage pool disorder.	Autosomal Dominant
<i>NBEAL2</i>	Gray platelet syndrome (GPS): macrothrombocytopenia with mild to moderate bleeding due to alpha granule deficiency, splenomegaly and bone marrow fibrosis	Autosomal Recessive
<i>P2RY12</i>	P2RY12-related platelet dysfunction (platelet-type bleeding disorder 8): mild-moderate mucocutaneous bleeding and excessive bleeding in response to trauma or surgery due to impaired platelet aggregation responses to ADP	Autosomal Recessive
<i>PLA2G4A</i>	PLA2G4A-related platelet dysfunction (cytosolic phospholipase-A2 alpha deficiency): platelet dysfunction from a metabolic defect and small bowel ulcers caused by decreased production of eicosanoids	Autosomal Recessive
<i>PLAU*</i>	Quebec Platelet Disorder (QPD): delayed onset bleeding, large trauma induced hematomas, hemarthrosis, muscle bleeds and hematuria from hyperfibrinolysis due to increased platelet urokinase plasminogen activator from a tandem 77.9kb duplication encompassing the PLAU gene	Autosomal Dominant
<i>PRKACG</i>	PRKACG-related thrombocytopenia (platelet-type bleeding disorder 19), characterized by severe macrothrombocytopenia with associated platelet dysfunction leading to moderate to severe bleeding	Autosomal Recessive
<i>RASGRP2</i>	RASGRP2-related platelet dysfunction (platelet-type bleeding disorder 18): moderate to severe bleeding and decreased platelet aggregation with ADP and epinephrine and in some cases arachidonic acid, collagen and thrombin	Autosomal Recessive
<i>RUNX1</i>	Familial platelet disorder with predisposition to myeloid leukemia (FPD/AML): mild to moderate thrombocytopenia with normal platelet size, platelet delta storage pool disorder and a predisposition to development of myeloid malignancies	Autosomal Dominant
<i>SLFN14</i>	SLFN14-related thrombocytopenia (platelet-type bleeding disorder 20): mild to moderate macrothrombocytopenia with associated platelet dysfunction from dense granule deficiency leading to variable bleeding	Autosomal Dominant
<i>SRC</i>	SRC-related thrombocytopenia (thrombocytopenia 6): thrombocytopenia and bleeding with associated myelofibrosis and bone pathology	Autosomal Dominant
<i>STIM1</i>	STIM1-related thrombocytopenia (Tubular aggregate myopathy and Stormorken syndrome): variable muscle weakness, miosis, thrombocytopenia with normal platelet size, hyposplenism, ichthyosis, dyslexia and short stature. Electron dense platelet inclusions and target-like organelles are characteristic	Autosomal Dominant
<i>TBXA2R</i>	Thromboxane receptor defect: pathogenic variants in TBXA2R have been proposed as contributing to a bleeding phenotype in the presence of additional pathogenic variants in genes affecting platelet function; these variants cause impaired platelet response to arachidonic acid and U46619 in vitro, but have not been shown to consistently correlate with a clinical phenotype	Risk allele
<i>TBXAS1</i>	TBXAS1-related platelet dysfunction (Ghosal syndrome; platelet-type bleeding disorder 14): increased bone density and platelet dysfunction due to impaired aggregation with arachidonic acid	Autosomal Recessive
<i>VIPAS39</i>	Arthrogryposis, renal dysfunction, and cholestasis syndrome type 2 (ARCS2): macrothrombocytopenia with platelet dysfunction from alpha granule deficiency with associated arthrogryposis, renal dysfunction, and cholestasis	Autosomal Recessive
<i>VPS33B</i>	Arthrogryposis, renal dysfunction, and cholestasis syndrome type 1 (ARCS1): macrothrombocytopenia with platelet dysfunction from alpha granule deficiency with associated arthrogryposis, renal dysfunction, and cholestasis	Autosomal Recessive

* Available only via aCGH

Indications for testing:

Platelet Function Disorder Panel (NGS and/or aCGH), order code 4835:

The Platelet Function Disorder Panel should be considered:

- In patients with lifelong bleeding compatible with platelet dysfunction without a diagnostic clinical or laboratory phenotype
- In patients with mild or variable thrombocytopenia with bleeding out of proportion to the degree of decrease in platelet counts raising concern for congenital platelet dysfunction
- In patients with thrombocytopenia and bleeding in whom platelet function testing cannot be obtained or is unreliable due to the severity of the thrombocytopenia

Single Gene Analysis (order code 4855) or Custom Blood Disorder Panel (Order Code 4850), (NGS and/or aCGH):

- Analysis of genes included in this panel may also be ordered as a standalone Single Gene Analysis or as a Custom Blood Disorder Panel (2-10 genes), by NGS and/or by aCGH, as dictated by the patient's clinical and laboratory phenotype, as well as their ancestry, or to supplement previous genetic testing.

Targeted Familial Variant Analysis (order code 4970):

Targeted variant analysis for clinical diagnosis, carrier identification, or prenatal diagnosis can also be performed on any gene in the panel when the pathogenic variant(s) is known in the family. If the proband was not tested at Versiti, a control sample is preferred and may be required (please call the laboratory to discuss). If the familial variant is a large deletion or duplication, aCGH for the involved gene is required.

For clinical questions about laboratory tests and test utilization support, contact Versiti Client Services: (414) 937-6396 or 800-245-3117, Option 1, to be directed to our genetic counselors and clinical support team.

Informed Consent

It is recommended that healthcare providers obtain informed consent from the patient when genetic testing is ordered, consistent with any applicable state laws and regulations, documenting that the patient has been advised of understands the indications for and implications of the genetic test. This panel is designed for clinical detection of germline genetic variants in genes with strong or definitive evidence for causality of platelet function disorders and/or inherited thrombocytopenia. Test results may nonetheless yield genetic findings that may be unrelated to the current clinical presentation, and/or may carry individual or familial implications such as risk for syndromic manifestation, predisposition to malignancy, and/or reproductive implications (such as carrier status). If needed, an informed consent form for Versiti Hematology Genetics testing can be found at <http://www.versiti.org/hg> under *forms*.

Test method:

NGS: This next-generation sequencing assay analyzes the complete coding region of 40 genes (excluding *PLAU*) plus a minimum 30bp of non-coding DNA, including intron-exon boundaries, and is compared to the build GRCh37.p13 reference sequence. *ANKRD26* analysis also includes approximately 200bp upstream of coding region to identify clinically significant variants in the 5'UTR. These targeted regions are captured by hybridization, amplified, and sequenced by massively parallel sequencing. Regions will have a minimum coverage of 50x and those regions with less than 50 sequencing reads or low quality coverage are supplemented with Sanger sequencing. All regions are covered by bidirectional analysis. Variants are identified by a customized bioinformatics pipeline, analyzed and comprehensively interpreted by our team of practicing hematologists with expertise in non-malignant hematology and laboratory diagnostics, scientists, and genetic counselors. All reported variants, including pathogenic, likely pathogenic, and variants of uncertain significance, are confirmed by Sanger sequencing. For prenatal testing, analysis of variable number tandem repeats (VNTR) is used to confirm results are not affected by maternal cell contamination.

aCGH: The specific genes are analyzed for copy number variations due to deletion or duplication by high density gene-focused array Comparative Genomic Hybridization. Probes are approximately 60bp in length and density of coverage in exonic regions is a minimum of 4 probes per 500 bp. Genomic DNA for the samples and gender-matched references are denatured, labeled with fluorescent dye and hybridized, the array is washed and scanned, and analysis is performed for the specific genes requested.

Assay sensitivity and limitations:

NGS: The analytical sensitivity of the NGS test is >99% for single nucleotide changes and insertions and deletions of less than 20 bp. NGS analysis is not designed to detect large deletions or duplications (>20 bp), or variants that are outside the regions sequenced. Low level mosaicism will not be detected by this sequencing methodology.

aCGH: Balanced chromosomal rearrangements (i.e., translocations, inversions) or point mutations that may be the cause of the clinical phenotype cannot be detected via aCGH. Any exonic deletion or duplication smaller than 500bp may not be detected. Low level of mosaicism will not be detected by aCGH. Probe performance could be affected by multiple SNPs in a given region. Breakpoints occurring outside the targeted gene(s) will not be defined.

Clinical Sensitivity

The clinical sensitivity of comprehensive genetic testing (NGS and aCGH) of the 41 genes in this panel is highest in patients with a history of lifelong bleeding suspected to be from inherited platelet dysfunction. In patients who

have thrombocytopenia and lacking or unclear platelet dysfunction, the Inherited Thrombocytopenia Panel or the Comprehensive Platelet Disorder Panel could be of higher diagnostic yield.

Reporting of Results

Results are classified and reported in accordance with ACMG next-generation sequencing and copy number variation standards and guidelines. Sequence variants and large deletions and duplications predicted to be pathogenic, likely pathogenic, and of uncertain significance will be reported; variants classified as likely benign or benign are typically not reported but such data are available upon request. Sequence variants are described using standard Human Genome Variation Society (HGVS) nomenclature (<http://hgvs.org>); copy number variants are described in accordance with the International System for Human Cytogenomic Nomenclature (ISCN).

Specimen Requirements

Parental/Patient/Pediatric: 3-5 mL Whole blood (EDTA tube, lavender top), 2-5 mL Bone marrow (EDTA tube, lavender top), 3-4 Buccal swabs, or $\geq 1\mu\text{g}$ of DNA at $\geq 50\text{ng}/\mu\text{L}$ of High Quality DNA.

Fetal: 7-15 mL amniotic fluid, 5-10 mg chorionic villi; back up culture of amniocytes or chorionic villi is highly recommended. Cultured: Two T25 flasks cultured amniocytes or chorionic villi (2×10^6 minimum). Maternal blood sample of 3-5 mL Whole blood (EDTA tube, lavender top) is requested for all prenatal samples for maternal cell contamination studies. For questions please contact the laboratory to discuss sample requirements.



ORDER

Required Forms

Please complete all pages of the requisition form. Clinical history (including patient's ethnicity, clinical diagnosis, family history, and relevant laboratory findings) is necessary for optimal interpretation of genetic test results and recommendations. Clinical and laboratory history can either be recorded on the

requisition form or clinical and laboratory reports can be submitted with the sample.

CPT Codes/Billing/Turnaround Time

Test code: 4835

For suggested CPT codes, visit the [Versiti.org/test menu](https://www.versiti.org/test-menu)

Turnaround time: 21 days

The CPT codes provided are subject to change as more information becomes available. CPT codes are provided only as guidance to assist clients with billing.

For additional information related to shipping, billing or pricing, please contact Versiti Client Services: (414) 937-6396 or 800-245-3117, Option 1, or LabInfo@versiti.org



SHIP

Shipping Requirements

Ship on an ice pack at room temperature. Protect from freezing. Place the specimen and the requisition into plastic bags and seal. Insert into a Styrofoam container, seal and place into a sturdy cardboard box, and tape securely. Ship the package in compliance with your overnight carrier guidelines. Label with the following address:

Client Services/Diagnostic Laboratory
Versiti
638 N. 18th St
Milwaukee, WI, 53233

References

Platelet Function Disorder references

1. Gresele P, Bury L, Falcinelli E. Inherited Platelet Function Disorders: Algorithms for Phenotypic and Genetic Investigation. *Semin Thromb Hemost.* 2016;42(3):292-305. doi:10.1055/s-0035-1570078
2. Lentaigne C, Freson K, Laffan MA, Turro E, Ouwehand WH; BRIDGE-BPD Consortium and the ThromboGenomics Consortium. Inherited platelet disorders: toward DNA-based diagnosis. *Blood.* 2016;127(23):2814-2823. doi:10.1182/blood-2016-03-378588
3. Megy K, Downes K, Simeoni I, et al. Curated disease-causing genes for bleeding, thrombotic, and platelet disorders: Communication from the SSC of the ISTH. *J Thromb Haemost.* 2019;17(8):1253-1260. doi:10.1111/jth.14479
4. Nurden AT, Nurden P. Inherited disorders of platelet function: selected updates. *J Thromb Haemost.* 2015;13 Suppl 1:S2-S9. doi:10.1111/jth.12898
5. Rand ML, Reddy EC, Israels SJ. Laboratory diagnosis of inherited platelet function disorders. *Transfus Apher Sci.* 2018;57(4):485-493. doi:10.1016/j.transci.2018.07.009
6. Rao AK. Inherited platelet function disorders: overview and disorders of granules, secretion, and signal transduction. *Hematol Oncol Clin North Am.* 2013;27(3):585-611. doi:10.1016/j.hoc.2013.02.005
7. Sharma R, Perez Botero J, Jobe SM. Congenital Disorders of Platelet Function and Number. *Pediatr Clin North Am.* 2018;65(3):561-578. doi:10.1016/j.pcl.2018.02.009
8. Songdej N, Rao AK. Inherited platelet dysfunction and hematopoietic transcription factor mutations. *Platelets.* 2017;28(1):20-26. doi:10.1080/09537104.2016.1203400

Variant interpretation references

9. Bean LJH, Funke B, Carlston CM, et al. Diagnostic gene sequencing panels: from design to report-a technical standard of the American College of Medical Genetics and Genomics (ACMG). *Genet Med.* 2020;22(3):453-461. doi:10.1038/s41436-019-0666-z
10. Rehm HL, Bale SJ, Bayrak-Toydemir P, et al. ACMG clinical laboratory standards for next-generation sequencing. *Genet Med.* 2013;15(9):733-747. doi:10.1038/gim.2013.92
11. Richards S, Aziz N, Bale S, et al. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. *Genet Med.* 2015;17(5):405-424. doi:10.1038/gim.2015.30
12. Riggs ER, Andersen EF, Cherry AM, et al. Technical standards for the interpretation and reporting of constitutional copy-number variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics (ACMG) and the Clinical Genome Resource (ClinGen). *Genet Med.* 2020;22(2):245-257. doi:10.1038/s41436-019-0686-8.